STAT718/BIOL703: Genomic Data Science Next Sequencing Data Analysis: Alignment (Chapter 5 in Gondro's book)

Dr. Yen-Yi Ho (hoyen@stat.sc.edu)

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Next Generation Sequencing (NGS) Data Analysis: Alignment

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- Alignment: BWT
- Simple DNA mapping
 - Bowtie2 and Samtools

R

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Work Flow



Mapping of Short Reads



Reference genome

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Tools in R



	Splice	Tool	Description	
1	No	BWA	Burrows-Wheeler Transform algorithm based	
			tool that accurately maps reads (up to 1 Mbp)	
			to a given reference genome.	
2	No	Bowtie2	Memory-efficient aligner for mapping very short	
			reads (ranging from 50 to 100bp) to large genomes.	
3	Yes	STAR	Spliced read aligner for de novo identification of	
			novel splice junctions. STAR is significantly faster	
			at read mapping compared with other	
			sequence aligners.	
4	Yes	Rsubread/	Uses Rbowtie but can detect spliced junction	
		QuasR		

Table: Commonly used tools for short read alignment

"Next generation sequencing technology and genomewide data analysis: Perspectives for retinal research", Progress in Retinal and Eye Research 55 (2016) 1e31.

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Alignment

Bowtie is a program that aligns short reads to an indexed genome. Burrows Wheeler transform (BWT) and the FM index are implemented by Bowtie to perform read alignments.

- the FM index: Ferragina, P. and Manzini, G. (2000), "Opportunistic data structures with applications"
- BWT: Burrows, M. and Wheeler, D.J. (1994), "A Block-sorting lossless data compression algorithm".

Sequence Mapping Algorithms



- Suffix Array
- Borrows-Wheeler transformation & LF mapping

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BW alignment

Seed

Break reference into k-mer words (seed) and hash their locations to speed later searches

<i>k-mer</i> : substring	Index of T		
of length k	CGTGC: 0,4		
	GCGTG: 3		
	GTGCC: 1		
	GTGCT: 5		
	TGCCT: 2		
	TGCTT: 6		

5-mer index

T: CGTGCGTGCTT

BLAST Algorithm Steps

- Seed-and-extend paradigm
- For each k-mer in query, find possible reference k-mers that matches well with it.
- Only words with $\geq T$ cutoff score are kept.
- For significant high-scoring segment pairs (HSP), use Smith-Waterman algorithm to join HSPs.



Sequence Mapping Algorithms



- Suffix Array
- Borrows-Wheeler transformation & LF mapping

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BW alignment

Computing BWT: An Example

$\mathsf{GCCACC} \to \mathsf{GCCACC}\$$

Then get rotation matrix (M):

\$GCCACC C\$GCCAC CC\$GCCA ACC\$GCC CACC\$GC CCACC\$G GCCACC\$

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Computing BWT: An Example

Then sort the rotation matrix.

F L \$GCCACC ACC\$GCC C\$GCCAC ↓CACC\$GC CC\$GCCA CC\$GCCA

We call the first column F and last column L. We can recover the whole sequence by storing only F and L. The index F and L can be stored in a very efficient manner (multiple Cs).

BWT: FM index

$$\begin{array}{cccc} \mathsf{F} & \mathsf{L} \\ \$ & C_0 \\ A_0 & C_1 \\ C_0 & C_2 \\ C_1 & C_3 \\ C_2 & A_0 \\ C_3 & C_0 \\ C_0 & \$ \end{array}$$

$$\overset{\leftarrow}{G_0C_3C_1A_0C_2C_0}$$

Alignment Algorithms

Before the alignment, it is important to know if the experiment was single-end or paired-end. The output of alignment step is commonly stored in SAM/BAM format.

- SAM file (.sam): Sequence Alignment/Map (SAM) is a text file that stores alignment information of reads to reference genome or given sequence. Some aligners such as STAR generate SAM file as an output of alignment process of short reads to reference genome. A SAM file includes a header section starting with @ character and alignment section consisting of multiple lines.
- BAM file (.bam): Binary Alignment/Map (BAM) is the binary version of SAM file. As SAM file does, BAM file stores alignment information of reads however BAM file is compressed (has smaller size) and more efficient in many sequencing analysis tools as it is compared to SAM file. A SAM file can be converted to a BAM file (or vice versa) with the help of SAMtools.

Headers	1 @HD VN:1.0 S0:unsorted	
Alignments	2 eSQ SR:gi1106402131ref1Nc_008253.11	INC_ (AAA 2222
	222222222222222222 A5:i:-3 XN:i:0 XM:i:1 X0:i:0 XG:i:0 NM:i:1 MD:Z:8665 YT:Z:UU 5 gil110640213 ref NC_008253.1 _31_476_0:0:0_0:0:0_1/1 16 gil110640213 ref 008253.1 407 42 70M * 0 0 GGAAAGCAATGCCAGGGCAGGGGCAGGTGGCCACCGTCCTC CCCCCGCCAAAATCACCAACCATCTG 22222222222222222222222222222222222	INC_ ICTG 2222 (T:Z
	<pre>G gil10640213 ref NC_008253.1 _210_743_2:0:0_1:1:0_2/1 0 gil10640213 ref 008253.1 210 42 70M * 0 0 CATTACCACCACCATCACCATTACCACGAGAAACGGTGCG GACGCCTACAGGAAACACCGAAAAAA 22222222222222222222</pre>	INC_ GGCT 2222 31A7

Each row describes a single alignment of a raw read against the reference genome. Each alignment has 11 mandatory fields, followed by any number of optional fields.

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DNA Alignment in R

There are R packages (Rsubread, and QuasR) available for implementing alignment in R. These program uses Rbowtie. (R code examples in Lab17.R)



RNA-seq alignment

- Transcriptome: the collection of all transcripts that can be generated from a genome.
- ► ≈ 92-94% of human transcripts with more than one exon have alternatively spliced isoforms. (Wang, Sandberg, Luo et al. Nature 2008) → the same gene generates multiple mRNA transcripts.
- Due to splicing, there is a difference in aligning a set of reads from an RNA sample and a DNA sample.

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RNA-seq alignment

There are a number of algorithms that are designed to align a set of short reads to transcriptome.

- STAR
- TopHat
- MapSplice
- RSubread (R)
- QuasR (R)

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R code using QuasR and RSubread are in Lab17.R. Linux command using STAR is also posted on the course website.

A list of software and comparison can be found at "Tools for mapping high-throughput sequencing data", Bioinformatics (2012) 28 (24): 3169-3177."

Work Flow



Getting Read Counts

There are several approaches to get read count from mapped RNA reads. The table below listed some of the popular approaches:

Function	Package	Framework	Output	DESeq2 input function
summarizeOverlaps	GenomicAligenments	R/Bioconductor	Summarized	DESeqDataSet
			Experiment	
featureCounts	Rsubread	R/Bioconductor	matrix	DESeqDataSetFromMatrix
tximport	tximport	R/Bioconductor	list of matrices	DESeqDataSetFromTximport
htseq-count	HTSeq	Python	files	DESeqDataSetFromHTSeq

We use summarizeOverlaps and featureCounts as examples in RNAseqAlignment2.R